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4-Methylpteridinones as orally active and selective PI3K/mTOR dual inhibitors

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ABSTRACT

Pteridinones were designed based on a non-selective kinase template. Because of the uniqueness of the PI3K and mTOR binding pockets, a methyl group was introduced to C-4 position of the pteridinone core to give compounds with excellent selectivity for PI3K and mTOR. This series of compounds were further optimized to improve their potency against PI3K α and mTOR. Finally, orally active compounds with improved solubility and robust in vivo efficacy in tumor growth inhibition were identified as well.

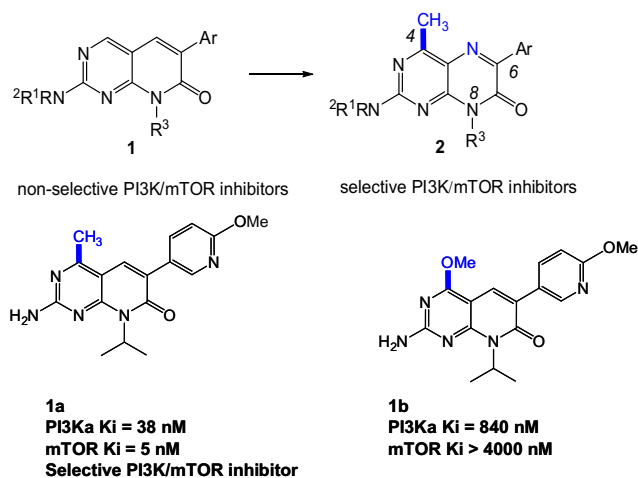
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The phosphatidylinositol-3-kinase (PI3K)/mammalian target of rapamycin (mTOR) signaling pathway plays a central role in driving tumor cell proliferation, survival, angiogenesis and metastasis by activating mutation, deletion or amplification on one or several of its components.¹ PI3Ks are lipid kinases that regulate cellular growth and metabolism by phosphorylation of the 3-hydroxy of phosphatidylinositol to generate phosphatidylinositol triphosphate (PIP₃). One of the pivotal downstream effectors, Akt, is activated by PIP₃ via membrane recruitment and phosphorylation at Thr-308 by the 3-phosphoinositide-dependent protein kinase-1 (PDK-1). The PIP₃ formation is also responsible for facilitating the second phosphorylation at Ser473 of Akt by mTOR–riCTOR complex (mTORC2).² Among different subtype of PI3Ks, PI3K α is the primary target for the treatment of cancers since functional loss of PTEN (phosphatase and tensin homolog protein is encoded by PTEN gene, the most commonly mutated tumor-suppressor gene in cancer after p53), oncogenic mutations in the PIK3CA gene encoding PI3K α , amplification of the PIK3CA gene and over-expression of Akt have been established in many human cancers.³

mTOR is a mammalian serine/threonine kinase and a member of PI3K like kinase (PIKK) family of proteins. Rapamycin and its analogs (e.g., RAD001 (everolimus), CCI-779 (temsirolimus), and AP23573 (deforolimus)) have demonstrated anti-tumor activity in clinical trials.⁴ Temsirolimus was approved in May 2007 for the treatment of advanced kidney cancer.⁵ mTOR exists in two dif-

ferent complexes,⁶ mTORC1, a rapamycin sensitive complex signaling to S6K1 and 4E-BP1 and mTORC2, an rapamycin insensitive complex signals to Akt as mentioned earlier. The existence of this rapamycin insensitive component of the mTOR signaling pathway thus provides new opportunities to inhibit mTOR.

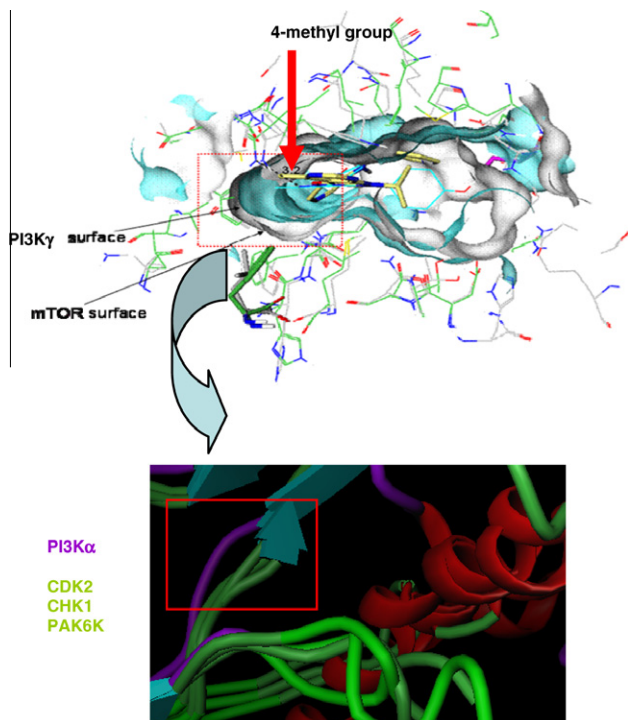
A small molecule inhibitor that targets mTOR kinase should prevent signaling through both mTORC1 and mTORC2 to have a



Scheme 1. From non-selective to selective PI3K/mTOR inhibitors.

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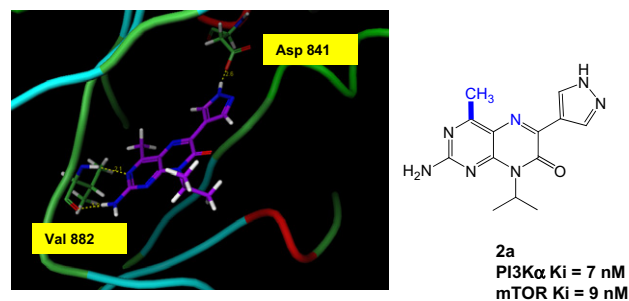
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Scheme 2. Top: the 4-methyl analog of compounds **1**, **1a**, was docking to PI3K γ and mTOR (PI3K surface: light blue, mTOR surface: light gray); bottom: PI3K α ATP binding site (purple color) vs protein kinases (green color: CDK2, CHK1 and PAK6K).

broad and advantageous spectrum of pharmacology over mTORC1 specific rapamycin. Given the pivotal role of the PI3K α /Akt/mTOR pathway in the biology of human cancers, we believe a dual PI3K α /mTOR inhibitor can provide a great understanding in cancer treatment.

Finding leads for this PI3K/mTOR dual inhibitor project, we started with promiscuous kinase hits with a general structure as compound **1** in Scheme 1. To improve compound selectivity against other kinases, PI3K ATP binding site was analyzed and compared with other protein kinase ATP binding sites. It is interesting to find out that there is an additional small binding pocket in the PI3Ks and it is not available in regular protein kinases as shown in Scheme 2. This additional binding pocket should be able to accommodate a larger molecule and give us the selectivity that



Scheme 3. Co-crystal structure of compound **2a** and PI3K γ .

we were looking for. From molecular modeling and docking studies, a small methyl group at C-4 position of compound **1** such as compound **1a** will orient the C-4 methyl group to fill this unique PI3K/mTOR binding pocket as shown in the top graph of Scheme 2. Compound **1a**⁷ was made to test this model and indeed this compound shows excellent selectivity against other kinases in our kinase screening panels which covers more than sixty different kinases. The compound also has good and moderate binding affinity on PI3K α and mTOR, respectively. This unique binding pocket for PI3K/mTOR turns out to be very small as we had predicted earlier since compound **1b** which has 4-methoxy instead of 4-methyl at C-4 position does not bind to mTOR at >4 μ M and has much weaker binding affinity for PI3K α as well.

To further increase structural diversity from **1**, we decided to replace the pyridopyrimidinone with pteridinone as the core structure (compound **2** in Scheme 1) while keeping the critical C-4 methyl group for selectivity. Based on earlier SAR learning from compound **1**,⁷ we jump started to make compound **2a** (Scheme 3) with the pteridinone core and hoped the SAR transfer well between these two series. Indeed, compound **2a** shows good potency on PI3K α and mTOR with great selectivity against other kinases as compound **1a**. We then focused on this 4-methyl-pteridinone series to improve both PI3K and mTOR potency with acceptable ADME properties.

To improve PI3K α and mTOR potency, small alkyl groups at R¹ and R² are preferred. From the co-crystal structure of compound **2a** and PI3K γ ⁸ (Scheme 3, PDB: 3OAW), it is consistent with our docking model and indicated that the C-2 amino and N-3 interact with Val 882 in the hinge region. Additionally, the pyrazole group at C-6 position picks up an additional interaction with Asp 841 (Scheme 3). Both Val 882 and Asp 841 are conserved in PI3K α as

Table 1
Representative compounds **2**, **2b–g** on the C-6 aryl modifications

<p>2b PI3K Ki = 20 nM mTOR Ki = 6 nM</p>	<p>2c PI3K Ki = 60 nM mTOR Ki = 113 nM</p>	<p>2d PI3K Ki = 82 nM mTOR Ki = 3940 nM</p>
<p>2e PI3K Ki = 6 nM mTOR Ki = 30 nM</p>	<p>2f PI3K Ki = 2 nM mTOR Ki = 18 nM</p>	<p>2g PI3K Ki = 27 nM mTOR Ki = 974 nM</p>

Table 2
Representative compounds **2**, **2h–l**

	<p>2h PI3K Ki: 2.75 nM mTOR Ki: 5.11 nM S473 pAKT IC₅₀: 5.74 nM HLM ER <0.3 RLM ER <0.3 DLM ER <0.3 RRCK = 21</p>		<p>2i PI3K Ki: 46 nM mTOR Ki: 0.8 nM S473 pAKT IC₅₀: 68 nM HLM ER <0.2 RLM ER = 0.5 DLM ER = 0.2</p>
	<p>2j PI3K Ki: 2.8 nM mTOR Ki: 6.8 nM S473 pAKT IC₅₀: 14.3 nM HLM ER <0.2 DLM ER = 0.2 RRCK = 32</p>		<p>2k PI3K Ki: 13 nM mTOR Ki: 5 nM S473 pAKT IC₅₀: 25 nM HLM ER <0.2 RLM ER = 0.3 DLM ER = 0.1</p>
	<p>2l PI3K Ki: 2.8 nM mTOR Ki: 0.85 nM S473 pAKT IC₅₀: 5.7 nM HLM ER <0.2 RRCK = 32</p>		

Val 851 and Asp 810. However, there is some subtle SAR on mTOR and PI3K potency in the C-6 binding pocket. Small changes in the C-6 aryl group will have profound effects on either PI3K or mTOR potency such as examples shown in Table 1.

Nevertheless, all the compounds are still selective against most of other protein kinases.

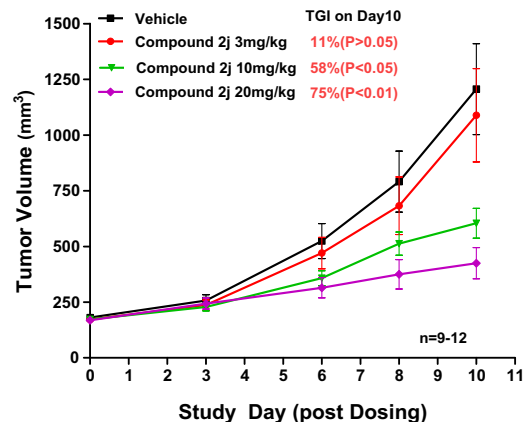
One ADME drawback in this series is solubility, and fortunately the R³ (N-8 position) points to the solvent exposed surface area according to co-crystal structures. Therefore, we can modify R³ to modulate ADME properties of pteridinones to improve compounds' solubility.

After mixing and matching, compounds **2h–l** were made with solvating groups at R³ for better solubility, and with promising heteroaryl groups at C-6 to interact with Asp 810 residue in PI3K α for better potency. All the compounds **2h–l** still keep their excellent selective against other kinases as **1a**. Compounds **2h–l** are potent binders for both PI3K α and mTOR with K_i <15 nM, and all the compounds are potent in our cellular functional assay, phosphorylation of S473 of AKT in BT20 cell line. In addition, these compounds have improved solubility and excellent microsomal stability across different species as shown in Table 2.

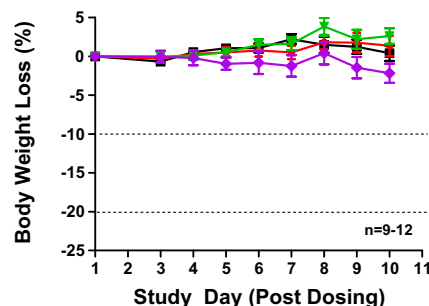
Compound **2j** was picked for our in vivo tumor growth inhibition (TGI) studies because of favorable human PK predictions. As shown in Scheme 4, a nice dose–response curve was observed in the TGI studies when **2j** was orally dosed in U87 (glioma cell line) mouse xenograft model. Additionally, there was no effect on body weight in the same studies.

General synthesis of compound **2** is described below and as shown on Scheme 5. Compounds of general structure **4** are prepared from 2,4-dichloro-6-methyl-5-nitropyrimidine (**3**) with ammonia or optionally substituted amines in THF and triethylamine as the base to yield compound **4**. Compound **4** without purification are then treated with ammonia or optionally substituted amines under the same condition as the first N-alkylation with or without triethylamine base to generate compound **5** in 40–

Antitumor activity of Compound **2j** in U87 Xenograft model

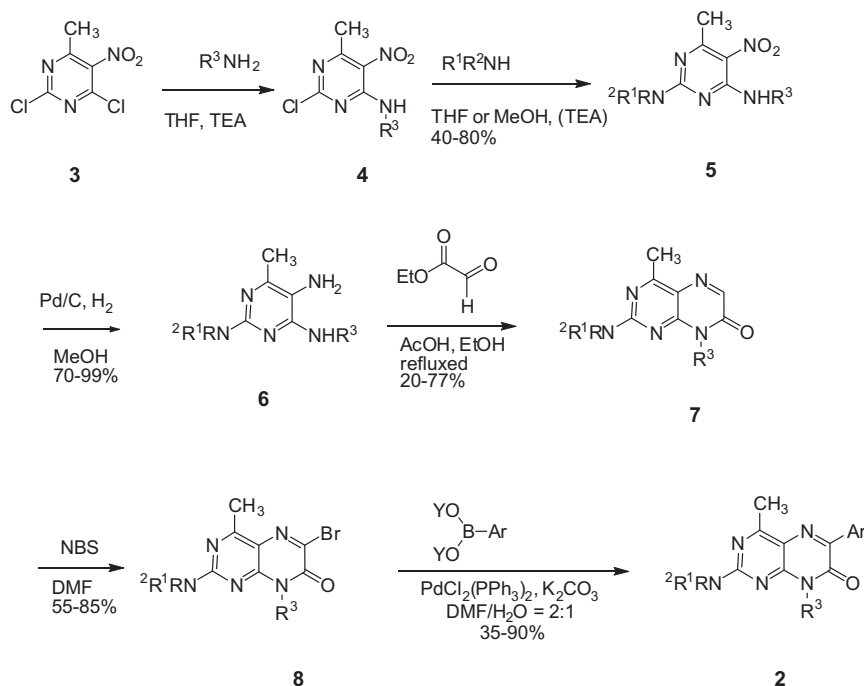


Effect on body weight of Compound **2j** in U87 xenograft model



Scheme 4. TGI and body weight studies of compound **2j**.

80% yield in two steps. The nitro group in compound **5** is reduced with Pd/C under hydrogenation in MeOH to give compound **6** with



Scheme 5. Synthesis of compounds 2.

free amino group in 70–99% yield. Compound **6** are cyclized with ethyl glyoxalate and acetic acid in refluxing ethanol to give compound **7** in 20–77% yield which are subsequently treated with NBS in DMF to yield brominated compound **8** in 55–85% yield. Finally, compound **6** are reacted with an appropriate aryl boronic acid or ester under the Suzuki–Miyaura coupling⁹ condition with catalytic amount of $\text{PdCl}_2(\text{PPh}_3)_2$ and 2 equiv of K_2CO_3 in the $\text{DMF}/\text{H}_2\text{O} = 2:1$ mixed solvent system at 100°C to generate compound **2** in 35–90% yield.

In summary, from a non-selective kinase template, 2-aminopyridopyrimidinone (**1**), 4-methylpteridinones were designed based on a small special pocket within PI3K and mTOR binding site to give us a series of selective PI3K/mTOR dual inhibitors. After optimization in compound potency and solubility, Compounds with good cellular potency were found. In addition, several orally active compounds were identified with robust efficacy in our tumor growth inhibition models as well.

References and notes

- Engelman, J. A.; Luo, J.; Cantley, L. C. *Nat. Rev. Genet.* **2006**, 7, 606.
- Sarbassov, D. D.; Guertin, D. A.; Ali, S. M.; Sabatini, D. M. *Science* **2005**, 307, 1098.
- (a) Broderick, D. K.; Chunhui, D.; Parrett, T. J.; Samuels, Y. R.; Cummins, J. M.; McLendon, R. E.; Fults, D. W.; Velculescu, V. E.; Bigner, D. D.; Yan, H. *Cancer Res.* **2004**, 64, 5048; (b) Samuels, Y.; Wang, Z.; Bardelli, A.; Silliman, N.; Ptak, J.; Szabo, S.; Yan, H.; Gazdar, A.; Powell, S. M.; Riggins, G. J.; Willson, J. K. V.; Sanford, M.; Kinzler, K. W.; Bert, V.; Velculescu, V. E. *Science* **2004**, 324, 554.
- Huang, S.; Houghton, P. J. *Curr. Opin. Pharmacol.* **2003**, 3, 371.
- Hudes, G.; Carducci, M.; Tomczak, P.; Dutcher, J.; Figlin, R.; Kapoor, A.; Staroslawaska, E.; Sosman, J.; McDermott, D.; Bodrogi, L. *N. Engl. J. Med.* **2007**, 356, 2271.
- Sarbassov, D. D.; Ali, S. M.; Sabatini, D. M. *Curr. Opin. Cell Biol.* **2005**, 17, 596.
- (a) Cheng, H.; Bhumralkar, D.; Dress, K.; Hoffman, J. E.; Johnson, M. C.; Kania, R. S.; Le, P. T. Q.; Nambu, M. D.; Pairish, M. A.; Plewe, M. B.; Tran, K. T. 2008, WO 2008032162.; (b) Bruendl, M. M.; Gogliotti, R. D.; Goodman, A. P.; Reichard, G. 2005 WO 2005105801.
- We cannot get PI3K α co-crystal structures so we used PI3K γ enzyme instead of PI3K α enzyme for our dual inhibitor program.
- A review on Suzuki–Miyaura reaction: Miyaura, N.; Suzuki, A. *Chem. Rev.* **1995**, 95, 2457.